

# Biochemical and histological studies on the possible protective impact of the herb basil (Ocimum basilicum) on adriamycin induced toxicity in rats. I. Influence on the liver.

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# Abstract

The purpose of this work was to evaluate the effect of basil (Ocimum basilicum) against hepatotoxicity induced in albino rats by the anticancer drug adriamycin (ADR). The biochemical results showed that adriamycin caused significant elevation in serum ALT (Alanine aminotransferase) and AST (Aspartate aminotransferase) enzymes after 4, 6 and 8 weeks of treatment. It also caused an increase in malondialdehyde (lipid peroxidation marker) and decrease in activities of the antioxidant enzymes, superoxide dismutase and catalase. This drug has resulted in various histological changes in the liver. These changes include impairment of the normal structural organization of the hepatic lobules, congestion and dilatation of blood vessels, cytoplasmic vacuolization of the hepatocytes, leucocytic infiltrations and fatty infiltration. Treating animals with ADR and basil (Ocimum basilicum) led to an improvement in both biochemical and histological changes induced by ADR. There are significant decreases in ALT and AST activity. Moreover, Ocimum basilicum reduced the level of malondialdehyde and increased the activity of superoxide dismutase and catalase. In conclusion the results of the present work indicated that Ocimum basilicum had a protective effect against liver damage induced by adriamycin and this is due to antioxidant activities of some substances found in water extract of Ocimum basilicum.

antoxidant enzyme, histology.

#### **1** Introduction

treatment course of cancer (Rizk et al., 2014). It is obtained Ocimum basilicum on the hepatotoxicity of the anticancer from a bacterial species (Streptomyces peucetius) and it drug, adriamycin in albino rats.

was introduced in 1969 for tumour treatment. It is an anthracycline antitumor antibiotic and has a highly effective role in many human tumours including certain types of bladder, lung, breast, ovarian and stomach cancer (Ogawa, 1985), Hodgkin's lymphoma (Hodgkin's disease), non-Hodgkin's lymphoma and acute leukemia (Blum and Carter, 1974). Its use is limited due to the development of hepatotoxicity, nephrotoxicity and cardiotoxicity (Tallaj et al, 2005). Hepatotoxicity is one of the main side effects associated with ADR treatment. Extensive investigations have been conducted on the hepatotoxicity as well as general organ toxicity of adriamycin (Deepa and Varalakshmi, 2003; El-Sayvad et al., 2009).

Human beings have probably used medicinal plants for thousands of years for cure of their various ailments and still continue to do so (Hill, 1989). These natural products have many biologic and pharmacologic properties (Hosseinimehr, 2014). Today, it is estimated that about 80% of the world population relies on botanical preparations as medicine to meet their health needs (Ogbera et al., 2010). Basil or sweet basil (Ocimum basilicum) is an important medicinal plant and a culinary herb widely cultivated in many countries that contains several antioxidants compounds and considered as a major volatile oil crop (Grayer et al., 1996). This plant is belonging to the family Lamiaceae (Ghosh, 1995). It is widely used in folk medicine to treat a wide range of diseases due to its numerous pharmacological activities. Many studies have Keywords: basil, adriamycin, liver, aminotransferases, reported that basil leaf extracts have potent antioxidant, anti-aging, anticancer, antiviral, and antimicrobial properties (Akujobi et al., 2004 ;Chiang et al., 2005 ; Bozin et al., 2006 ; Manosroi et al., 2006 ; Almeida et al., 2007). Adriamycin (Doxorubicin) is a drug used in the The present study was conducted to reveal the influence of

#### 2 Materials and Methods

#### Chemicals

Adriamycin (Doxorubicin): Doxorubicin (Adriablastina produced by Carlo Erba) was purchased from a local pharmacy in the form of 10 mg/ampoule.

Basil extract: A fresh leaves of Basil (Ocimum basilicum) was collected from the garden in Faculty of Science -Menoufia University, Shebin El- Kom, Egypt. The leaves were rinsed with clean water to remove any foreign matter. Leaves were blended with distilled water. The mixture was strained, the merc pressed and the mixture was filtrated using filter paper. The aqueous extract was used at a dose level of 20 ml/kg Ocimum basilicum (Offiah and Chikwendu, 1999).

### Animals and treatments

Healthy adult male albino rats (Rattus norvegicus) weighing  $120 \pm 5$  g were used. They were obtained from the breeding center of experimental animals, Helwan, Egypt. Animals were kept in the laboratory under constant temperature  $(24\pm2^{\circ}C)$  for at least one week before and throughout the experimental work. They were maintained on a standard diet composed of 55% corn starch, 20% casein, 15% corn oil, 5% salt mixture and 5% vitaminzed starch (Egyptian Company of Oils and Soap Kafr-El Zayat, Egypt). Water was available ad libitum. All the experiments were done in compliance with the guide for the care and use of laboratory animals (National Research Council, 1985).

Animals were divided into 4 groups:

Group 1: These animals (20 rats) were served as normal controls

Group 2: These animals (20 rats) were treated with oral aqueous O. basilicum extract at a dose level of 20 mg/kg for 5 days/wk through 8 weeks.

Group 3: Animals of this group (20 rats) were injected intraperitonealy with ADR at a dose level of 2 mg/kg body weight in sterile saline, once per week for 8 weeks

Group 4: Animals of this group (20 rats) treated with ADR at the same dose level as those of group 3 followed by oral administration aqueous O. basilicum extract at a dose level of 20 mg/kg for 5 days/wk through 8 weeks.

#### **Biochemical assays**

For biochemical assays blood was collected and centrifuged at 3000 rpm for 10 minutes and stored at - 20 °C. Liver function enzymes, ALT and AST, were determined in serum according to the method of Gella et al. (1985). malondialdehyde was determined by the method of Ohkawa et al. (1979) and superoxide dismutase was assayed using the method of Rest and Spitznagel (1977). Catalase activity in the sera was determined according to the method of Aebi et al. (1974).

# **Histological preparations**

Immediately after decapitation, the animals were dissected, their livers were removed from treated and control animal groups and fixed in 10% formalin. After Fig. 1: Effect of Adriamycin and/or basil on body weights fixation, the specimens were dehydrated using an ascending series of alcohol, cleared in 2 changes of xylene, and

embedded in molten paraffin (melting point: 50-58°C). Sections of 5 mm thickness were cut using rotary microtome and mounted on clean slides. For histopathological examination, the sections were stained with Ehrlich hematoxylin and counter stained with eosin.

### Statistical analysis

The data were expressed as mean  $\pm$  standard error. Data were analyzed using Student's t-test and homogeneity of variances (Levene test) using statistical program of social science (SPSS) software for windows. P < 0.05, P < 0.01and P < 0.001 values were used.

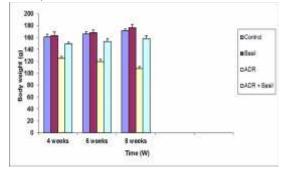
### **3 Results**

#### Change in total body weight

Data in (Fig. 1) showed that treating animals with ADR for 8 weeks caused significant decrease in body weight of rats compared with control group. On the other hand, administration of O. basilicum and ADR caused significant increase in body weight of rats compared with ADR group. When O. basilicum was given to animals, insignificant change in body weight was recorded.

# **Biochemical results**

Treatment with ADR for 8 weeks caused a highly significant elevation (P < 0.001) in the activity of ALT and AST as compared to those of the control animals. On the other hand, treatment with ADR and O. basilicum extract showed a reduction in the activity of these two enzymes (Figs. 2, 3). Both control and animals given O. basilicum showed no significant differences in serum activity of ALT and AST. Lipid peroxidation marker (MDA) increased significantly in ADR-treated animals when compared to the control group. Animals treated with ADR and O. basilicum showed a significant decrease in MDA when compared with ADR group (Fig. 4). Data expressed in (Fig. 5) and (Fig. 6) show that there are no significant differences in levels of SOD and CAT in both control and O.basilicum extract groups. Animals treated with ADR revealed that levels of SOD and CAT decreased significantly (P < 0.001) after 4, 6 and 8 weeks of treatment. On the other hand, treating animals with ADR and O. basilicum extract led to an increase in these antioxidant enzymes. This increase was highly significant (P < 0.001) after 8 weeks of treatments.



of rats.

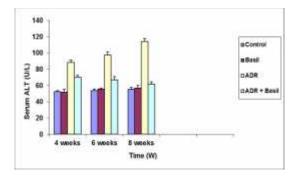


Fig. 2: Effect of Adriamycin and/or basil on serum ALT.

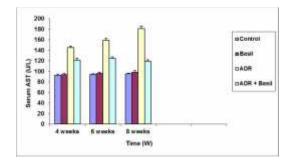


Fig. 3: Effect of Adriamycin and/or basil on serum AST.

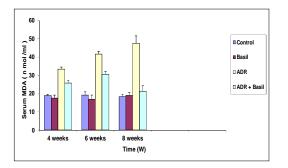


Fig. 4: Effect of Adriamycin and/or basil on serum MDA

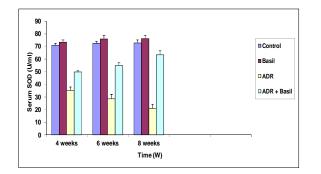


Fig. 5: Effect of Adriamycin and/or basil on serum SOD

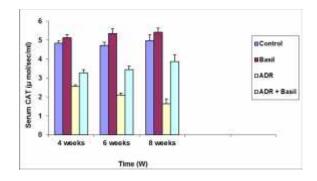


Fig. 6: Effect of Adriamycin and/or basil on serum CAT

## **Histological results**

Liver of control rat showed normal lobular architecture. The hepatic cells were found arranged in strands around the central vein and sinusoids appeared containing Kupffer cells (Fig. 7). Liver obtained from rats treated with basil extract exhibited the normal structure (Fig. 8). Whereas animals treated with ADR for 4 weeks revealed that the hepatic tissue was injured and the hepatic blood vessels were enlarged and congested (Fig. 9). After 6 weeks, the hepatic tissue lost its normal organization and Most of the hepatocytes showed cytoplasmic vacuolation with pyknotic nuclei (Fig. 10). After 8 weeks of treatment, these changes became intensive. The hepatic architecture was lost, clearly fatty infiltrations and masses of leukocytic infiltration were observed (Fig. 11, 12). After treatment with both ADR and O. basilicum, an improvement was recorded in the hepatic tissue. In these specimens, the hepatocytes appeared normal with an increase of binucleated cells (Fig. 13).

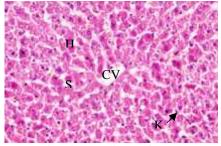


Fig. 7: Section in the liver of a control rat showing the basic structure: central vein (CV), hepatocytes (H), blood sinusoids (S) and Kupffer cells (K) (X400).

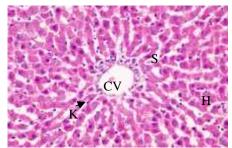


Fig. 8: Section in the liver of a rat treated with basil extract showing the hepatic architecture appeared normal, where the hepatic cells (H) arranged around central vein (CV) in

observed (X400).

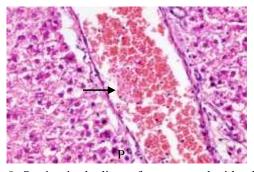


Fig. 9: Section in the liver of a rat treated with adriamycin for 4 weeks showing congested and dilated blood vessel Fig. 13: Section in the liver of a rat treated with ADR and pyknotic nuclei (P) (X400).

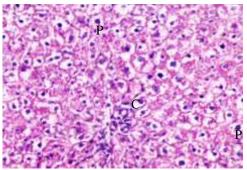


Fig. 10:Section in the liver of a rat treated with adriamycin for 6 weeks showing loss of the characteristic hepatic architecture, cytoplasmic vaculation of the hepatocytes (C) and pyknotic nuclei (P) (X400).

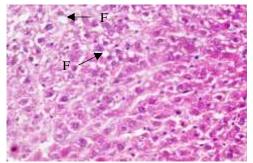
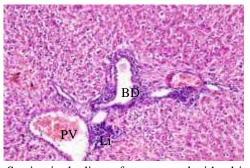
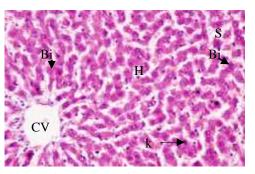


Fig. 11: Section in the liver of a rat treated with adriamycin for 8 weeks showing loss of the characteristic hepatic architecture and fatty infiltration (F). (X400).



for 8weeks showing congested and enlarged portal vein increased the MDA levels and decreased SOD and CAT

cords, Blood sinusoids (S) and Kupffer cells (K) were also (PV), enlarged and proliferated bile ductule (BD)and leucocytic infiltration (Li)



(arrow), disruption in hepatocytes architecture with O. basilicum showing advanced degree of improvement, restoration of structure of the liver tissue and binucleated cell (Bi) is observed (X400).

#### **4** Discussion

Adriamycin is a very important agent in the treatment of cancer patients although its use may be complicated by the presence of acute and chronic side effects. It has been shown that free radicals are involved in adriamycin - induced toxicities (Yagmurca et al., 2004). It has been reported that adriamycin caused severe damage in some organs like liver, testes, heart and kidneys (Shivakumar et al., 2012). Adriamycin was found to cause the generation of free radicals and the induction of oxidative stress that correlates with cellular injury (Saad et al., 2001). The present results revealed that ADR exerted deleterious action on the liver of rats both biochemically and histologically. Among the most sensitive and widely used liver enzymes are the aminotransferases. They include aminotransferase alanine (ALT) and aspartate aminotransferase (AST). These enzymes are normally predominantly contained within liver cells and to a lesser degree in the muscle cells. If the liver is injured or damaged, the liver cells spill these enzymes into the blood, raising the AST and ALT enzyme blood levels and signaling liver disease. Adriamycin was found to affect serum transaminases (ALT, AST). Significant increase in ALT and AST levels of sera of ADR treated rats was recorded in the present study. In agreement with this result Roomi et al. (2014) found that administration of ADR to old male BALB/c mice resulted in a marked increase of hepatic ALT and AST. Several investigators also obtained similar results. In this concern, Ito et al. (2001) observed increase in the levels of ALT and AST in the serum of adriamycin administered rats. Similarly, Guo et al. (2016) reported that administration of ADR at dose of 20 mg/kg increases the levels of ALT and AST. Swamy et al. (2011) proved that administration of ADR for two weeks elevated the activity of serum marker enzymes ALT and AST. A high lipid peroxidation with a decrease in the antioxidant enzymes, superoxide dismutase (SOD) and catalase (CAT) were recorded in liver of adriamycin-treated rats. Similarly, Fig. 12: Section in the liver of a rat treated with adriamycin Swamy et al. (2012) observed that administration of ADR

study investigated that male Wister rats given a single i.p. (intraperitonial) injection of 15mg/kg/body weight of ADR caused increased the MDA levels and decreased SOD and CAT levels (Al-Sowayan and Mahmoud, 2014). Koti et al. (2009) recorded an increase in the levels of MDA and decrease in the levels of SOD and CAT after administration ADR. Many histological alterations were observed in liver of ADR treated rats. The most marked signs of tissue impairment were congestion of blood vessels, leucocytic infiltrations, cytoplasmic vaculation of the hepatocytes and fatty infiltrations. In agreement with these results, Gokcimen et al. (2007) reported that ADR increased mononuclear cells infiltration, congestion of blood vessels and necrosis. Marked changes in liver of male albino rats such as congestion of blood vessels, leucocytic infiltrations and fatty infiltration after treatment with ADR was observed by Sakr and Abo-El-Yazid (2012). Abbas (2011) protection may be also due to the anti-inflammatory noticed changes in hepatic tissue including inflammatory property of ocimum which reduces formation, release, and cell infiltration, necrosis and vacuolation after ADR activity of inflammatory mediators such as cytokines, administration. Also, Taher et al. (2013) revealed that histamine, prostaglandins, and leukotrienes. Ocimum higher doses of ADR caused massive hepatotoxicity treatment was found to exhibit hepatoprotective effect as including dissolution of the hepatic cords, focal recorded by enhanced activity of the antioxidant enzyme, inflammation, apoptosis and necrosis of the hepatic tissues SOD and CAT and diminished amount of lipid with fibrosis around the portal area. Lower doses exhibited abnormal changes including vacoulation of the hepatocytes with widening of the sinusoidal capillaries in addition to congestion and vasodilatation of the central veins.

The present findings demonstrated that *O.basilicum* improves the biochemical and histological changes induced in the liver by ADR. This indicated the effectiveness of O.basilicum in prevention of ADR hepatotoxicity. In diazinon and O. basilicum extract caused a reduction of agreement with these observations the hepatoprotective MDA level and elevation of both SOD and CAT activity. effects of O.basilicum have been shown in many studies on Kath and Gupta (2006) showed that hydroalcoholic extract experimental liver damage. Yamamoto et al. (2005) proved of ocimum sanctum leaves lowered levels of MDA and that *O.basilicum* suppressed hepatic fibrosis and protected increased levels of SOD in animal models of peptic ulcer. liver against parenchymal damage induced by CCl<sub>4</sub>. The aqueous extract of Ocimum americanum leaves at doses of 200 and 400 mg /kg p.o. has significant hepatoprotective ability against paracetamol - induced hepatic damage in rats. The levels of some serum biochemical parameters such as ALT and AST in rats intoxicated with paracetamol alone had their serum AST and ALT levels significantly increased (Bayomy and Salah Eldin, 2015). The pretreatment of rats with aqueous extract of O. americanum leaves caused a significant decrease in the serum levels of ALT and AST (Aluko et al., 2013). Oral doses of basil 5 References extract of 200 mg/k g b.w. significantly decreased the higher levels of serum of ALT and AST induced by CCl<sub>4</sub> (Yacout et al., 2012). Lahon and Das (2011) reported that sanctum have hepatoprotective effect against О. paracetamol toxicity in rats. The hepatoprotective effects of O.basilicum have been shown in studies on experimental liver damage. Sakr et al. (2011) reported that treating animals with CCl<sub>4</sub> and aqueous leaf extracts of O. basilica led to an improvement, in both histopathological and biochemical alterations induced by CCl<sub>4</sub>. A study by Arhoghro et al. (2012) indicated that aqueous leaf extracts of O. gratissimum has anti-hepatotoxic action against

levels compared to the level of the normal group. Also, a cisplatin induced hepatic toxicity in rats. The ameliorating effect was further evident through marked decrease in the activities of the marker enzymes ALT and AST and decreased histopathological alterations of liver tissues. Mahboub and Arisha (2015) showed that treating rats with diazinon and O. basilicum extract improved the histological structure of the liver and caused significant decrease of ALT and AST. These results suggested that O. basilicum protects the hepatocytes from injuries and improves liver function. Rasekh et al. (2012) decided the safety of administration and found that Ocimum basilicum microscopic examination of liver in control and treated animals with O. basilicum showed no toxic effects on this organ and supported safety of O. basilicum data was inferred from biochemical parameters. Chinnasamy et al., (2007) reported that the protective action of ocimum was attributed to its antioxidant action. They added that this peroxidation against the adriamycin-induced hepatotoxicity in rats. These results are in agreement with other studies which proved the antioxidant effects of O. basilicum extract. Sakr and Abdel samie (2015) reported that lipid peroxidation marker, MDA, was increased while the antioxidant enzymes namely SOD and CAT were decreased after Diazinon treatment. Treating animals with Ramesh and Satakopan (2010) reported that administration of Ocimum sanctum extract before and after cadmium intoxication resulted in a significant decrease in lipid peroxidation (LPO) levels and significant increase in SOD and CAT levels. Therefore, the results of the present work indicate that O. basilicum has protective effect against hepatoxicity induced by adriamycin and this is may be attributed to its antioxidant activities of some substances found in water extract of Ocimum basilicum.

Abbas, E.K. (2011):Histopathological changes caused by doxorubicin in laboratory mice (Mus Musculus L). J Pak Med Assoc. 61(11):1108-10.

Aebi, H.; Wyss, S.R.; Scherz, B. and Skvaril, F. (1974):Heterogeneity of erythrocyte catalase II. Isolation andcharacterization of normal and variant erythrocyte catalase andtheir subunit. Enzyme, 17: 307-318.

Akujobi, C.O.; Anyanwu, B.N.; Onyeze, G.O. and Ibekwe, V.I. (2004): Antibacterial activities and preliminary phytochemical screening of four medicinal plants. J.Appl. Sci., 7(3): 4328-4338.

Al-Sowayan, N. and Mahmoud, N.H. (2014): The doxorubicin hepatotoxicity. Hum. Exp. Toxicol., 26(6): Protective Effect of Grape Seed Extract on Cardiotoxicity Induced by Doxorubicin Drug in Male Rats. Advances in Bioscience and Biotechnology, 5(12):1078-1089.

Almeida, I.; Alviano, D.S., Vieira, (2007): Antigiardial activity of Ocimum basilicum essential tochemistry 43, 1041 - 1047. oil. Parasitol. Res, 101 (2): 443-452.

(2013): Hepatoprotective activity of Ocimum americanum L leaves against paracetamol - induced liver damage in with sesamin via the suppression of oxidative stress. Hum rats,. American Journal of Life Sciences, 1(2): 37-42.

Arhoghro, E. M.; Ikeh, C.; Uwakwe, A. A.; Ekpo, K. E. and Anosike, E. O. (2012): Curative potential of Aqueous extract of scent leaf (Ocimum gratissimum) on Environ. Health., 29(4):341-53 cisplatin induced hepatotoxicity in albino wister rats. Journal of Pharmaceutical and Scientific Innovation, 1(4):1-8.

Bayomy, M.F.F and Salah Eldin, A.A. (2015): Ameliorative influence of ginko biloba extract on acetaminophen-induced oxidative stress in livers and kidneys of male albino rats.J Bio Sci. Appl. Res., 1:91-96.

Blum, R.H. and Carter, S.K.(1974): Adriamycin. A new anticancer drug with significant clinical activity. Ann Intern Med.80(2):249-59.

Bozin, B.; Mimica-Dukic, N.; Simin, N. and Anackov, G. (2006): Characterization of the volatile composition of essential oils of some lamiaceae spices and the antimicrobial and antioxidant activities of the entire oils. J. Agric. Food Chem., 54 (5): 1822-1828.

Chiang, L.C.; Ng, L.T.; Cheng P.W.; Chiang W. and Lin, C.C. (2005): Antiviral activities of extracts and selected pure constituents of Ocimum basilicum.Clin. Exp. Pharmacol. Physiol., 32(10):811-816.

Chinnasamy, S.; Regini, G.; Kingston, C.; Kavitha, A.; Sukumaran, S. and Raj, A.(2007): Potential Antiinflammatory Properties of CrudeAlcoholic Extract of Ocimum basilicum L. in Human Peripheral Blood Mononuclear Cells.J. Of Health Science, 53: 500-505.

Deepa, P.R. and Varalakshmi, P. (2003): Protective Lett. 235 (1): 114-120. effect of low molecularweight heparin on oxidative injury and cellular abnormalities inadriamycin-induced cardiac Hepatoprotective effect of Ocimum basilicum extract and hepatic toxicity. Chemico. Biol.Interact., 146(2): 201-210.

El-Sayyad, H.I.; Ismail, M.F.; Shalaby, F.M.; Abou-El-Magd, R.F.; Gaur, R.L.; Fernando, A.; Raj, M.H. and Ouhtit, A. (2009): Histopathological effects of cisplatin, doxorubicin and 5-flurouracil (5-FU) on the liver of malealbino rats Int. J. Biol. Sci., 5(5): 466-473

Moreno, R.; Durban, R. and Gomez, J.A. (1985): A simple procedure for routine determination of aspartate 330. aminotransferase and alanine aminotransferase with pyridoxal phosphate. Clin. Chem. Acta., 153: 241-247.

Ghosh, G.R. (1995): Tulasi (N.O. Labiatae, Genus-Ocimum) .New Approaches to Medicine and Health (NAMAH), 3:23-29

Gokcimen, A.; Cim, A.; Tola, H.T.; Bayram, D.; stomach cancer. Kitakant Med. J., 35: 463-476. Kocak, A.; Ozgüner, F. and Ayata, A. (2007): Protective effect of N-acetylcysteine, caffeic acid and vitaminE on

519-525.

Grayer R.; Bryan S.; Veitch N.; Goldstone F.; Paton A. and Wollenweber E. (1996):External flavones from D.P. sweet basil, Ocimum basilicumand related taxa. Phy-

Guo, H.; Liu, Y.; Wang, L.; Zhang, G.; Su, S.; Zhang, Aluko, B. T.; Oloyede, O. I. and Afolayan, A. J. R.; Zhang, J.; Li, A.; Shang, C.; Bi, B. and Li, Z.(2016): Alleviation of doxorubicin-induced hepatorenal toxicities Exp Toxicol. [Epub ahead of print]

> Hosseinimehr, S.J.( 2014): Beneficial effects of natural products on cells during ionizing radiation. Rev.

> Hill, A.F.(1989): Economic Botany: A Text Book of Useful Plants and Plant Products. 2nd Edn. New York: McGraw Hill Book Company, Inc.

> Ito, K.; Ozasa, H.; Nagashima, Y.; Hagiwara, K. and Horikawa S. (2001): Pharmacological preconditioning with doxorubicin, Implications of heme oxygenase-1 induction in doxorubicin-induced hepatic injury in rats, Biochem Pharmacol, 62:1249.

> Kath, R.K. and Gupta, R.K.(2006): Antioxidant activity of Hydroalcoholic leaf extract of Ocimum sanctum in animal models of peptic ulcer. Indian J Physiol Pharmacol., 50(4): 391-396.

> Koti B.C., Vishwanathswamy, A.H.; Wagawade, J. and Thippeswamy, A.H (2009): Cardioprotective effect of lipistat against doxorubicin induced myocardial toxicity in albino rats. Indian Journal of Experimental Biology., 47:41-46

> Lahon, K. and Das, S. (2011): Hepatoprotective activity of Ocimum sanctum alcoholic leaf extract against paracetamol-induced Albino liver damage in rats.Pharmacognosy Res., 3(1):13-8.

> Manosroi, J.; Dhumtanom, P. and Manosroi A.(2006): Anti-proliferative activity of essential oil extracted from Thai medicinal plants on KB and P388 cell lines. Cancer

> Mahboub, Arisha, F.A and S.M. (2015): against the toxicity of diazinon in albino rats: Histopathological and immunohistochemical evaluation World J Pharm Sci., 3(5): 790-799.

> National Rsearch Council (1985). Guide for use and care of laboratoryanimals, Publication no. 85-23, Washington, NIH.

Offiah, V.N. and Chikwendu, U.A. (1999): Gella, F.J.; Olivella, T.; Cruz-Pastor, M.; Arenas, J.; Antidiarrhoeal effects of Ocimum gratissimum leaf extract in experimental animals. J Ethnopharmacol., 68(1-3):327-

> Ohkawa, H.; Ohishi, N. and Yagi, K. (1979): Assay for lipid peroxides in animals tissues by thiobarbituric acid reaction. Anal. Boi. Chem., 95(2): 351-358.

> Ogawa T (1985): Studies on endoscopic local injection of an anticancer agent into experimental canine

Ogbera, A.O.; Dada, O.; Adeyeye, F. and Jewo, P.I. (2010): Complementary and alternative medicine use in B.C. and Manjula, D.V.(2012): Cardioprotective effect of diabetes mellitus. West Afr. J. Med. 29(3): 158-162.

Taher, M.T.; Al-Sammak, M.A. and AL-Qzazz, M.M. (2013): Effect of Doxorubicin on the Histological Structure of the Liver in Male Albino Rats J Med J., 47 (3):220-226. (2012):Hepatoprotective effect of basil (Ocimum basilicum

Tallaj, J.A.; Franco, V.; Rayburn, B.K.; Pinderski, L.; Benza, R.L.; Pamboukian, S.; Foley, B. and Bourge, R.C.(2005): Response of doxorubicin induced cardiomyopathy to the current management strategy of Ucar, M. and Fadillioglu, E.(2004): Caffeic acid phenethyl heart failure. J Heart Lung Transplant., 2005;24(12):2196-2201.

Ramesh, B. and Satakopan, V. N. (2010): Antioxidant 34. Activities of Hydroalcoholic Extract of Ocimum sanctum Against Cadmium Induced Toxicity in Rats. Indian J Clin T. Biochem., 25(3): 307-310.

Rasekh, H.R.; Hosseinzadeh, L.; Mehri, S.; Nejad, M.K .; Aslani, M. and Tanbakoosazan, F.(2012): Safety Assessment of Ocimum Basilicum Hydroalcoholic Extract in Wistar Rats: Acute and Subchronic Toxicity Studies.Iran J Basic Med Sci. 15(1): 645-653.

Rest, R.F. and Spitznagel, J.K. (1977): Subcellular distribution of superoxide dismutases in human neutrophils. Influence of myeloperoxidase on the measurement of superoxide dismutase activity. Biochem. J., 166: 145-153.

Rizk. S.M.; Zaki, H.F. and Mina, M.A.M. (2014): Propolis Attenuates Doxorubicin-Induced Testicular Toxicity in Rats Food and Chemical Toxicology., 67: 176-186.

Roomi, M.W.; Kalinovsky, T.; Roomi, N.W.; Rath, M. and Niedzwieck, A. (2014): Prevention of Adriamycininduced hepatic and renal toxicity in male BALB/c mice by a nutrient mixture Exp Ther Med., 7(4): 1040–1044.

Saad, S.Y.; Najjar, T.A. and Al-Rikabi, A.C. (2001): The preventive role of deferoxamine against acute doxorubicin-induced cardiac, renal and hepatic toxicity in rats.Pharmacol.Res., 43: 211-218.

Sakr, S.A.; El-Abd, S.F.; Osman, M.; Kandil, A.M. and Helmy, M.S.(2011) Ameliorative Effect of Aqueous Leave Extract of Ocimum Basilicum on Ccl4 - Induced Hepatotoxicity and Apoptosis in Albino Rats. Journal of American Science 7(8): 116-127.

Sakr, S.A., Abo-El-Yazid, S.M. (2012): Effect of fenugreek adriamycin-induced seed extract on hepatotoxicity and oxidative stress in albino rats. Toxicol Ind Health.,28(10):876-85.

Sakr, S.A and Abdel Samie, H.A. (2015): Protective effect of Ocimum basilicum leaves extract on diazinon induced reproductive toxicity and oxidative stress in albino rats. Ejpmr., 3(1), 375-384.

Shivakumar, P.M.; Rani, M.U.; Reddy.; A.G. and Anjaneyulu, Y. (2012): A Study on the Toxic Effects of Doxorubicin on the Histology of Certain Organs Toxicol Int. 19(3): 241–244.

Swamy, A.V.; Wangikar, U.; Koti, B.C.; Thippeswamy, A.H.; Ronad, P.M. and Manjula, D.V.(2011): Cardioprotective effect of ascorbic acid on doxorubicin-induced myocardial toxicity in rats.Indian J Pharmacol.43(5): 507-511.

Swamy, A.V.; Gulliaya, S. Thippeswamy, A. Koti, curcumin against doxorubicin-induced myocardial toxicity in albino rats: indian J Pharmacol., 44(1): 73-77.

Yacout, G.A.; Elguindy, N.M and El Azab, E.F. L.) on CCl4-induced liver fibrosis in rats African Journal of Biotechnology 11(90):15702-15711.

Yagmurcaa, M.; Erdogan, H.; Iraz, M.; Songur, A.; ester as a protective agent against doxorubicin nephrotoxicity in rats.Clinica Chimica Acta, 348(1-2): 27-

Yamamoto, J.; Yamada, K.; Naemura, A.; Yamashita, and Arai, R.(2005): Testingvarious herbs for antithrombotic effect.Nutrition., 21:580-587.