

Characteristics of peripheral Leukocyte in moderate infection of COVID-19

Ahmed Abdelhalim Yameny

Society of Pathological Biochemistry and Hematology, Egypt

***Corresponding Author:** Ahmed A. Yameny **E-mail:** dr.ahmedyameny@yahoo.com Tel: (002)01222708665, (002)01002112248

DOI: 10.21608/jbaar.2021.251237

Abstract:

Background: Blood leukocytes are an important part of the body's defense system, and infection status can be predicted by measuring WBC levels COVID-19 may involve many organ systems in its host. Studies suggest that hematological profiles change during SARS-CoV-2 illness. **Patients and methods:** This study included 504 mild infected patients with confirmed COVID-19 infection, these study subjects were randomly selected irrespective of the age group and both genders were included, EDTA blood sample was collected for performing total and differential white blood cells (Diagon D-cell 60 hematology analyzer Europe-Diagon Ltd. Hungary). **Results:** The present study included patients aged from14 years to 75 years mean age was 44.5 ± 30.5 who were confirmed to have Covid-19 based on real-time reverse transcription-polymerase chain reaction, female gender was more frequent (n=280, 55.6%) than Male gender (n=224, 44.4%). This study reveals normal total WBCs count in 320 patients (63.5%), neutrophilia with a sensitivity of 77.8% and lymphopenia has a sensitivity of 73% for diagnosis or prognosis of mild infection of COVID-19 patients (Outpatients and patients under home observation).

Keywords: COVID-19, WBCs, neutrophilia, lymphopenia, biomarker.

1. Introduction:

The coronavirus disease 2019 (COVID-19) outbreak began in early December 2019 in Wuhan, China⁽¹⁾, the WHO officially named the disease caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) as COVID-19 and then officially declared the COVID-19 outbreak a pandemic on March 11, 2020 (WHO, 2020)⁽²⁾.

The previous study mainly focused on the immune dysfunction caused by severe acute respiratory

syndrome coronavirus (SARS-CoV) and Middle East respiratory syndrome coronavirus (MERS-CoV), respectively. Coronavirus infections (SARS and MERS) are confirmed to activate both innate and adaptive immune responses ^(3,4). Simply it means that the changes in peripheral blood cells could reflect the immune damage caused by virus infection.

In mild cases, immune responses were efficiently established to curb the viral replication, while in severe cases, uncontrolled inflammation and microcirculation dysfunctions together lead to viral sepsis with immunologic impairment ⁽⁵⁾. As components of blood routine tests available for almost all hospitalized patients, PBICs usually serve as practical markers in infectious diseases. Indeed, leukocytosis, leukopenia, and lymphopenia have been reported to be commonly seen in COVID-19 patients ⁽⁶⁾.

Blood leukocytes are an important part of the body's defense system, and infection status can be predicted by measuring WBC levels ⁽⁷⁾. COVID-19 may involve many organ systems in its host. Studies suggest that hematological profiles change during SARS-CoV-2 illness. Neutrophils are involved in early anti-viral defense. However, during severe pneumonia, neutrophils become cytotoxic through degranulation and lysis ⁽⁸⁾. Studies have suggested that neutrophil recruitment may exacerbate COVID-19 immunopathology ⁽⁹⁾.

Lymphocytes play a crucial role in maintaining immune homeostasis during virus infection, especially SARS-CoV-2⁽¹⁰⁾, among hematological parameters, lymphopenia is associated with disease severity; patients who have died from COVID-19 have had significantly lower lymphocyte counts than survivors. Repletion of lymphocytes may be an important factor for recovery⁽¹¹⁾ (Henry, 2020).

Thus, the differentiation of peripheral white blood cells may indicate the immunologic impairment at the early stage of the disease. So, this study was performed to assess the value of peripheral total white blood cells, neutrophils, and lymphocyte cells in mild infection of COVID-19 patients, this study included outpatients and patients under home observation, while hospitalized patients are not included in this study.

2. Patients and methods

2.1. Study population Patients

This study included 504 hospitalized patients with confirmed COVID-19 infection, these study subjects

were randomly selected irrespective of the age group and both genders were included.

It was performed following the ethical standards laid down in the 1964 Declaration of Helsinki and its later amendments. All the studied population was informed about the purpose of sample collection. Patients were free to refuse to include in this study.

2.2. Data collection

In this cross-sectional study, we obtained data regarding 504 hospitalized patients with confirmed COVID-19 via real-time reverse transcription-polymerase chain reaction (PCR), they came to Alyameny laboratory in Alexandria, Egypt for biomarkers and complete blood count investigations, and reviewed the medical records and compiled data between August 12 and December 30, 2020.

2.3. Collection and processing of blood samples:

EDTA blood sample was collected for performing complete blood count and platelet count (Diagon D-cell 60 hematology analyzer Europe-Diagon Ltd. Hungary) on 504 Positive COVID-19 patients for individuals matching in age and gender.

2.4. Assay procedure as manufactory instructions:

Diagon D-cell 60 hematology analyzer Europe-Diagon Ltd. Hungary was used for white blood cell count, considered normal WBCs count: $4 - 10 \times 10^{9}$ /L, normal neutrophils: 45- 70%, and normal lymphocytes: 20-45%

2.5. Statistical analysis

Data were analyzed using SPSS statistical software, version 20.0(SPSS, Chicago, Illinois, USA). All continuous data are presented as means and standard deviations, while categorical data are presented as numbers and percentages. A sensitivity analysis was performed to analyze relationships between COVID-19 infected patients and WBCs, neutrophils, and lymphocytes count.

3. Results:

The present study included patients aged from 14 years to 75 years mean age was 44.5 ± 30.5 who were confirmed to have Covid-19 based on real-time reverse transcription-polymerase chain reaction, the female gender was more frequent (n=280, 55.6%) than the Male gender (n=224, 44.4%).

Table (1): shows the percentage of COVID-19 mild infected patients (out hospitalized and home observation Patients) concerning total white blood cells count, this study reveals normal WBCs count in 320 patients (63.5%), low WBCs count in 40 patients (7.9%), and high WBCs count in 144 patients (28.6%). Table (2): shows the percentage of COVID-19 mild infected patients (out hospitalized and home observation Patients) concerning neutrophils count, This study reveals normal neutrophils count in 112 patients (22.2%), there were no infected patients with low neutrophils count, and high neutrophils count in 392 patients(77.8%), then the high-level of neutrophils has a sensitivity of 77.8%, it was a significant biomarker for COVID-19 diagnosis or prognosis in out-hospitalized patients (Outpatients and patients under home observation).

Table (3): shows The percentage of COVID-19 mild infected patients (out hospitalized and home observation Patients) concerning lymphocytes count, This study reveals normal lymphocytes count in 112 patients (22.2%), low lymphocytes count in 368 patients(73%), and high lymphocytes count in 24 patients(4.8%), then the low-level of lymphocytes has a sensitivity of 73%, it was a significant biomarker for COVID-19 diagnosis or prognosis in out-hospitalized patients (Outpatients and patients under home observation).

| | WBCs count | | | | | | | |
|----------------------------------|------------|-----|--------|------|------------|------|-------|------|
| COVID-19 POSITIVE Patients | Low level | | Normal | | High-level | | Total | |
| | No. | % | No. | % | No. | % | No. | % |
| Male | - | - | 168 | 52.5 | 56 | 38.9 | 224 | 44.4 |
| Female | 40 | 100 | 152 | 47.5 | 88 | 61.1 | 280 | 55.6 |
| Total | 40 | | 320 | | 144 | | 504 | |

Table (1): Association between WBCs count and mild infection of COVID-19 Patients

Normal WBCs count 63.5%, low 7.9%, and high 28.5%

| | Neutrophils count | | | | | | | |
|----------------------------------|-------------------|---|--------|----|------------|------|-------|------|
| COVID-19 POSITIVE Patients | Low level | | Normal | | High-level | | Total | |
| | No. | % | No. | % | No. | % | No. | % |
| Male | - | - | 56 | 50 | 168 | 42.9 | 224 | 44.4 |
| Female | - | - | 56 | 50 | 224 | 57.1 | 280 | 55.6 |
| Total | | | 112 | | 392 | | 504 | |

Table (2): Association between neutrophils count and mild infection of COVID-19 Patients

High neutrophils count 77.8 %

Table (3): Association between lymphocytes count and mild infection of COVID-19 Patients

| | Lymphocytes count | | | | | | | |
|----------------------------------|-------------------|------|--------|------|------------|-----|-------|------|
| COVID-19 POSITIVE Patients | Low level | | Normal | | High-level | | Total | |
| | No. | % | No. | % | No. | % | No. | % |
| Male | 152 | 41.3 | 72 | 64.3 | - | - | 224 | 44.4 |
| Female | 216 | 58.7 | 40 | 35.7 | 24 | 100 | 280 | 55.6 |
| Total | 368 | | 112 | | 24 | | 504 | |

Low lymphocytes count 73%

4. Discussion:

COVID-19 is an acute infectious disease caused by SARS-CoV-2. Most patients experience clinical symptoms, but some patients have no symptoms and are known as asymptomatic patients, the hematological changes may reflect a homeostatic mechanism to prevent systemic over-activation of inflammation.

This study was conducted on 504 mild infected COVID-19 patients (out hospitalized and home observation Patients) who were confirmed to have Covid-19 based on real-time reverse transcriptionpolymerase chain reaction, this study reveals a normal count for monocyte cells, eosinophil cells, and monocyte cells. But total WBCs were normal in 320 patients (63.5%), low WBCs count in 40 patients (7.9%), and high WBCs count in 144 patients (28.6%), Gao et al., 2020 reported that total white blood cell count was normal in all 43 cases (28 milds, 15 severe) ⁽¹²⁾, also Li et al., reported WBCs count was normal in all 54 cases ⁽¹³⁾, another study (meta-analysis) by Li et al., (2020b) on 1994 cases showed low WBCs count in 29% of cases⁽¹⁴⁾, Huang et al., 2020 reported high WBCs count in all 13 ICU cases⁽¹⁵⁾.

This study showed normal leukocyte count at 63.5%, leukopenia at 7.9%, leukocytosis at 28.6%, while neutrophilia has a sensitivity of 77.8% and lymphopenia has a sensitivity of 73% in mild COVID-19 infection which was less than CRP has a sensitivity 90.2%⁽¹⁶⁾, while other biomarkers have different sensitivity in mild COVID-19 infection as ferritin which has a sensitivity of 71.4%⁽¹⁷⁾, LDH has a sensitivity 67.7%⁽¹⁸⁾, and D-dimer has a sensitivity $36.4\%^{(19)}$.

Neutrophils are immune cells that are well known to be present in various lung diseases, including viral respiratory disease ⁽²⁰⁾. As a hallmark of the pathophysiology, it is widely accepted that neutrophils can exit the circulation into the airways, either through the postcapillary venule in the systemic circulation or through the capillary in the pulmonary circulation ^(21,22), this study showed high neutrophils count or neutrophilia has a sensitivity of 77.8% as a biomarker for COVID-19 diagnosis or prognosis in mild infected COVID-19 patients (Outpatients and patients under home observation).

Neutrophils are involved in early anti-viral defense. However, during severe pneumonia, neutrophils become

cytotoxic through degranulation and lysis ⁽²³⁾, Huang et al., 2020 reported neutrophilia in 13 ICU cases ⁽¹⁵⁾, and Wu et al., (2020) reported neutrophilia in 201 ARDS cases⁽²⁴⁾, but a study by Liu et al., showed low neutrophils count in most cases ⁽¹³⁾.

Several cohort studies have reported that lymphopenia can predict prognosis in COVID-19 patients ^(25,26).

A study involving 90 hospitalized COVID-19 patients reported an association between lymphopenia and disease severity⁽²⁷⁾, this study showed lymphopenia has a sensitivity of 73%, and it was a significant biomarker for COVID-19 diagnosis or prognosis in mild COVID- 19 patients (Outpatients and patients under home observation).

Other studies showed Lymphopenia is a factor related to poor prognosis in disease development (Ruan et al., 2020; Yang et al., 2020)^(28,29).

Chen et al., 2020a reported lymphopenia in 69% of cases (15 mild's, 9 severe, 5 critical)⁽³⁰⁾, while Huang et al., 2020 reported low lymphocyte count in ICU cases only⁽¹⁵⁾, Zhang et al., 2020 reported lymphopenia in all 140 infected cases⁽³¹⁾, another study (meta-analysis) by Li et al., (2020b) on 1994 cases showed lymphopenia in most cases⁽¹⁴⁾, Liu et al., 2020 also reported lymphopenia in most cases⁽¹³⁾, Mo et al., 2020 showed lymphopenia in all tested cases (70 mild's, 85 severe cases)⁽³²⁾,

The decreased production, apoptosis, and redistribution of lymphocytes may together lead to circulating lymphopenia⁽³³⁾.

Conflict of interest

There are no conflicts of interest.

Financial support and sponsorship

This research did not receive any specific grant from funding agencies in the public, commercial, or not-forprofit sectors.

5. References:

- Zhu, N., Zhang, D., Wang, W., Li, X., Yang, B., Song, J., et al. (2020). A Novel Coronavirus from Patients with Pneumonia in Chin. N. Engl. J. Med. 382,727–733. doi: 10.1056/NEJMoa2001017
- World Health Organization WHO. (2020). WHO Main Website. Available at: https://www.who.int (Accessed August 05, 2020).
- Yang M, Li CK, Li K, Hon KL, Ng MH, Chan PK, et al. Hematological findings in SARS patients and possible mechanisms (review). Int J Mol Med.2004; 14:311–5.
- 4. Chu H, Zhou J, Wong BH, Li C, Chan JF, Cheng ZS, et al. Middle east respiratory syndrome

coronavirus efficiently infects human primary T lymphocytes and activates the extrinsic and intrinsic apoptosis pathways. J Infect Dis. 2016; 213:904–14.

- Li H, Liu L, Zhang D, Xu J, Dai H, Tang N, et al. SARS-CoV-2 and viral sepsis: observations and hypotheses. Lancet. 2020; 395:1517–20.
- A.J. Rodriguez-Morales, J.A. Cardona-Ospina, E. Gutiérrez-Ocampo, et al., Clinical, laboratory and imaging features of COVID-19: A systematic review and meta-analysis, Travel. Med. Infect. Dis. (2020),

https://doi.org/10.1016/j.tmaid.2020.101623.

- Guan, W. J., Ni, Z. Y., Hu, Y., Liang, W. H., Ou, C. Q., He, J. X., et al. (2020). Clinical Characteristics of Coronavirus Disease 2019 in China. N. Engl. J. Med. 382, 1708–1720. doi: 10.1056/NEJMoa2002032
- Haick AK, Rzepka JP, Brandon E, Balemba OB, Miura TA. Neutrophils are needed for an effective immune response against pulmonary rat coronavirus infection, but also contribute to pathology. J Gen Virol 2014; 95:578e90.https://doi.org/10.1099/vir.0.061986-0.
- 9. Didangelos A. COVID-19 hyper inflammation: what about neutrophils? mSphere 2020;5(3). https://doi.org/10.1128/mSphere.00367-20.
- 10. Channappanavar R, Zhao J, Perlman S. T cellmediated immune response to respiratory coronaviruses. Immunol Res. 2014; 59:118–28.
- 11. Henry BM. COVID-19, ECMO, and lymphopenia: a word of caution. Lancet Respir Med; 2020.
- Gao Y, Li T, Han M, et al. Diagnostic Utility of Clinical Laboratory Data Determinations for Patients with the Severe COVID-19. J Med Virol. 2020.
- 13. Liu M, He P, Liu HG, et al. Clinical characteristics of 30 medical workers infected with new coronavirus pneumonia. Zhonghua Jie He He Hu Xi Za Zhi. 2020;43: E016.

- 14. Liu Y, Yang Y, Zhang C, et al. Clinical and biochemical indexes from 2019-nCoV infected patients linked to viral loads and lung injury. Sci China Life Sci. 2020b; 63:364–74.
- Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan. China. Lancet. 2020; 395:497–506.
- Sabah Farouk Alabd and Ahmed Abdelhalim Yameny. C-Reactive Protein as a Prognostic Indicator in COVID-19 mild infection Patients. *Journal of Medical and Life Science*, 2021; 3(2): 38-43. doi: 10.21608/jmals.2021.240126
- Ahmed Abdelhalim Yameny. Ferritin as a biomarker of infection in COVID-19 nonhospitalized patients. J Biosci App Res .2021;7(1): 23-28. doi: 10.21608/jbaar.2021.172371
- Ahmed Abdelhalim Yameny. Lactate dehydrogenase level as a COVID-19 biomarker. J Biosci App Res .2021;7(1): 29- 34. DOI21608/jbaar.2021.173662: 10.
- Ahmed Abdelhalim Yameny. D-dimer levels in COVID-19 out-hospitalized patients in Egypt. Journal of medical and life science.2021;3(1): 19-24. DOI: 10.21608/JMALS.2021.200216
- J.V. Camp, C.B. Jonsson, A role for neutrophils in viral respiratory disease, Front. Immunol. 8 (2017), <u>https://doi.org/10.3389/fimmu.2017.00550</u>.
- A. Juliana, R. Zonneveld, F.B. Plötz, et al., Neutrophil-endothelial interactions in respiratory syncytial virus bronchiolitis: An understudied aspect with a potential for prediction of severity of disease, J. Clin. Virol. 123 (2020), https://doi.org/10.1016/j.jcv.2019.104258.
- J. Gane, R. Stockley, Mechanisms of neutrophil transmigration across the vascular endothelium in COPD, Thorax 67 (2012) 553–561.
- Haick AK, Rzepka JP, Brandon E, Balemba OB, Miura TA. Neutrophils are needed for an effective immune response against pulmonary rat

221

coronavirus infection, but also contribute to pathology. J Gen Virol 2014; 95:578e90.https://doi.org/10.1099/vir.0.061986-0.

- 24. Wu C, Chen X, Cai Y, et al. Risk Factors Associated with Acute Respiratory Distress Syndrome and Death in Patients with Coronavirus Disease 2019 Pneumonia in Wuhan. China: JAMA Intern Med; 2020.
- 25. Qin C, Zhou L, Hu Z, Zhang S, Yang S, Tao Y, et al. Dysregulation of immune response in patients with coronavirus 2019 (COVID-19) in Wuhan, China. Clin Infect Dis. 2020; 71:762–8.
- 26. Li D, Chen Y, Liu H, Jia Y, Li F, Wang W, et al. Immune dysfunction leads to mortality and organ injury in patients with COVID-19 in China: insights from ERS-COVID-19 study. Signal Transduct Target Ther. 2020; 5:62.
- 27. Tan L, Wang Q, Zhang D, et al. Lymphopenia predicts disease severity of COVID-19: a descriptive and predictive study. Sig Transduct Target Ther2020; 5:33. https://doi.org/10.1038/s41392-020-0148-4.
- Ruan, Q., Yang, K., Wang, W., Jiang, L., and Song,
 J. (2020). Correction to: Clinical Predictors of Mortality Due to COVID-19 Based on an Analysis

of Data of 150 Patients from Wuhan, China. Intensive Care Med. 46, 1294–1297.doi: 10.1007/s00134-020-06028-z

- Yang, X., Yu, Y., Xu, J., Shu, H., Xia, J. A., Liu, H., et al. (2020). Clinical Course and Outcomes of Critically Ill Patients With SARS-CoV-2 Pneumonia in Wuhan, China: A Single-Centered, Retrospective, Observational Study. Lancet Respir.Med. 8, 475–481. doi: 10.1016/S2213-2600(20)30079-5
- Chen L, Liu HG, Liu W, et al. Analysis of clinical features of 29 patients with 2019 novel coronavirus pneumonia. Zhonghua Jie He He Hu Xi Za Zhi. 2020; 43:203–8.
- Zhang JJ, Dong X, Cao YY, et al. Clinical characteristics of 140 patients infected with SARS-CoV-2 in Wuhan. China: Allergy; 2020.
- 32. Mo P, Xing Y, Xiao Y, et al. Clinical characteristics of refractory COVID-19 pneumonia in Wuhan. China. Clin Infect Dis. 2020.
- 33. Tan L, Wang Q, Zhang D, Ding J, Huang Q, Tang YQ, et al. Correction: Lymphopenia predicts disease severity of COVID-19: a descriptive and predictive study. Signal Transduct Target Ther. 2020; 5:61.